

International Journal for Parasitology 30 (2000) 1453-1457



www.parasitology-online.com

Research note

Differential influence of *Pomphorhynchus laevis* (Acanthocephala) on the behaviour of native and invader gammarid species

Alexandre Bauer, Sandrine Trouvé, Arnaud Grégoire, Loïc Bollache, Frank Cézilly*

Laboratoire Ecologie-Evolution (UMR CNRS 5561 Biogéosciences), Université de Bourgogne, 6 bd. Gabriel, 21000 Dijon, France Received 18 August 2000; received in revised form 13 September 2000; accepted 13 September 2000

Abstract

Although various species of acanthocephalan parasites can increase the vulnerability of their amphipod intermediate hosts to predation, particularly by altering their photophobic behaviour, their influence on the structure of amphipod communities and the success of invader species has so far received little attention. We compared the prevalence and behavioural influence of a fish acanthocephalan parasite, *Pomphorhynchus laevis*, in two species of amphipods, *Gammarus pulex* and *Gammarus roeseli* in sympatry in the river Ouche (Burgundy, eastern France). There, *G. pulex* is a resident species, whereas *G. roeseli* is a recent coloniser. Both uninfected *G. pulex* and *G. roeseli* were strongly photophobic, although less so in the invading species. However, there was no significant difference in reaction to light between infected and uninfected *G. roeseli*, whereas infected *G. pulex* were strongly photophilic. We discuss our results in relation to the parasite's ability to manipulate invading host species, the possibility that resistant individuals have been selected during the invasion process, and the role that acanthocephalan parasites can play in shaping the structure of amphipod communities. © 2000 Australian Society for Parasitology Inc. Published by Elsevier Science Ltd. All rights reserved.

Keywords: Host-parasite relationship; Behavioural modifications; Favourisation; Biological invasions; Freshwater Crustacea; Gammarus pulex; Gammarus roeseli

In various freshwater systems in western Europe, populations of native amphipods are progressively displaced by introduced amphipods [1-3] that can show explosive range extension [4,5]. Several mechanisms, such as differential environmental tolerance [6], sterile interspecific mating [7], or differential aggression and mutual predation [3,8,9] can be involved in the regulation of the competitive interactions between native and introduced amphipod species. Additionally, both theoretical [10-12] and empirical evidence [13-15] have shown that parasites can play a major role in structuring animal communities. Recently, some studies have emphasized the importance of parasitism for the dynamics of amphipod populations and communities [16,17]. In particular, the outcome of biological invasions can depend to a large extent on the mediation of competition between invading and native species by parasites, i.e. 'apparent' competition [18]. On the one hand, the transmission of parasites from invading hosts to naive native hosts can mediate the displacement of the native species [19,20]. On the other hand, invasion can be facilitated if the invading

Various species of acanthocephalan parasites are exploiting amphipods as intermediate hosts. All acanthocephalan parasites rely on trophic transmission to complete their life cycle [22], and some species have evolved the ability to alter the behaviour of their intermediate host [23] to make it more vulnerable to predation by their definitive hosts, i.e. favourisation [24]. Interactions between acanthocephalan parasites and their amphipod hosts have been widely studied [25–27], whereas the influence of acanthocephalan parasites on the structure of amphipod communities and the success of invader species has received little attention. Recently, however, Dunn and Dick [17] observed that the prevalence of a bird acanthocephalan, Polymorphus minutus, was higher in the native amphipod Gammarus duebeni celticus than in the invader species Gammarus tigrinus in a freshwater site in Ireland.

Here we compare the behavioural alteration due to a fish acanthocephalan parasite, *Pomphorhynchus laevis*, in two species of amphipods, *Gammarus pulex* and *Gammarus roeseli* in sympatry. *Gammarus pulex* is a resident species in France, whereas *G. roeseli* is a recent coloniser of Central European origin [1,28,29]. The spreading of *G. roeseli* from

species is less susceptible than the native one to endemic parasites [21].

^{*} Corresponding author. Tel.: +33-38-0396246; fax: +33-38-0396231. *E-mail address:* fcezill@u-bourgogne.fr (F. Cézilly).

the Danube system to other rivers in western Europe seems to have been facilitated by the recent development of the canal network, especially in France [30]. The species has been reported to be infected with *P. laevis* in Czechoslovakia [31,32], while its susceptibility to the same parasite in other parts of its geographic range remains undocumented.

P. laevis is a non-specific parasite with larval stages (cystacanths) infecting various amphipod species (gammarids) whereas adults develop in several freshwater fishes [33]. The cystacanths of P. laevis are yellow-orange, and can be seen through the cuticle of infected gammarids that then show conspicuous yellow-orange dots. In addition to modified appearance, cystacanths of P. laevis are known to induce various changes in their intermediate host's phenotype. Infected gammarids show reduced O₂ consumption [34], increased haemocyanin concentration [35,36], and altered reaction towards light [26,27,37].

The study was conducted in the river Ouche at the Parc de la Colombière (Dijon, eastern France). In this site, G. roeseli lives in sympatry with infected G. pulex, with G. pulex being by large the more abundant species (approximately 90–95% G. pulex and 5–10% G. roeseli, G. Bauer, unpublished data). Gammarids were collected using the 'random-kick sampling' method [38], with a net retaining all size classes. We estimated the prevalence of G. laevis in G. pulex and G. roeseli from a large sample (G = 1087 from which there was 1036 G. pulex and 51 G. roeseli) collected in April 1999.

Several samples were collected in the same site (Ouche river) to obtain enough infected gammarids for behavioural experiments. In the laboratory, amphipods were maintained in aquaria filled with a mixture of water from the river and tap water (in a 50% proportion), maintained at 16°C, oxygenated and filtered. They were used in experiments within 72 h after collection.

A total of 459 gammarids were tested individually, to study the influence of species, sex, size and infection status on behaviour; amphipods harboring immature parasites were not retained in the analysis. At the end of the experiment, each amphipod was killed in 70% alcohol and sexed from the shape and size of segment 6 (propodus) of gnathopods 1 and 2, and the presence of eggs or embryos in the brood pouch. Individuals were measured by linear dimensions (body height at the level of the fourth coxal plate basis, see [39]). All gammarids were dissected for parasites. Parasite identity was determined following criteria of Brauer [40] and Brown et al. [41].

The experimental design and procedure used to quantify reaction to light is the same as described in Cézilly et al. [27]. Each individual was used only once in this setup. At the beginning of each trial, a single individual was placed at the bottom in the middle of the tank. After an acclimation period of 5 min, its position was recorded at 30 s intervals during 5 min. At the end of each trial, the behaviour of the individual was scored from the number of times that it was recorded in the light half of the aquarium. Thus, the score could range from 0 (strongly photophobic) to 10 (highly

photophilic). A score of 5 indicated no preference in relation to light. Because the distribution of scores did not conform to a normal distribution, data were analysed using non-parametric statistics [42,43]. Results were considered significant at P < 0.05. Two-tailed tests of significance are used throughout.

Overall, 1087 gammarids were collected in April 1999. The prevalence of *P. laevis* in the river Ouche did not differ between *G. pulex* (8.78%; n = 1036) and *G. roeseli* (7.84%; n = 51; Fisher's exact test: P = 1).

Overall, 194 *G. pulex* individuals (39 uninfected and 50 infected males, 35 uninfected and 70 infected females) and 265 *G. roeseli* individuals (60 uninfected and 38 infected males, 101 uninfected and 66 infected females) were used in the experiments.

We first considered the influence of parasite load on reaction to light. Infected G. pulex were assigned to three different categories corresponding to individuals harboring one (n = 63), two (n = 32) or three and more (n = 25) cystacanths. We found no effect of parasite load on reaction to light (Kruskal–Wallis: $H_2 = 2.720$, P = 0.26). Infected G. roeseli were assigned to two categories corresponding to individuals harboring one (n = 86) or two and more (n = 18) cystacanths. We found no effect of parasite load on reaction to light (Mann–Whitney U-test: Z = 1.657, P = 0.1). Therefore, for each gammarid species, data from all infected individuals were pooled in subsequent analyses.

Sex had no influence on reaction to light in either G. pulex (Mann-Whitney U-test, non-infected individuals: Z = -0.268, P = 0.788; infected individuals: Z = 0.749, P = 0.454) or G. roeseli (non-infected individuals: Z = 0.527, P = 0.60; infected individuals Z = -1.108, P = 0.27). To investigate if size influences reaction to light, we created three size classes, each one including about the same number of individuals. Because G. roeseli individuals are bigger than G. pulex ones, the groups were defined as presented in Table 1. Size has no effect on reaction to light in G. pulex (Kruskal-Wallis, healthy indivi- $H_2 = 0.733$, P = 0.69; infected individuals: $H_2 = 2.640$, P = 0.27) and in G. roeseli (Kruskal-Wallis, healthy individuals: $H_2 = 4.431$, P = 0.11; infected individuals: $H_2 = 1.348$, P = 0.51). Therefore, data for both sex and size were pooled for subsequent analysis.

Table 1 Number of gammarids (infected and uninfected) in each class size. Sizes correspond to the height of the metacoxal plate

Class size (mm)	Infected	Uninfected	
G. pulex			
1.843-2.315	26	27	
2.316-2.673	55	21	
2.674-3.845	39	26	
G. roeseli			
2.108-2.508	35	54	
2.509-2.853	46	54	
2.854-3.762	23	53	

There was a significant difference in reaction to light between infected and uninfected G. pulex and G. roeseli (Kruskal–Wallis analysis of variance: $H_3 = 48.295$, P < 0.00001). Among all categories, the only non-significant comparison was found between infected and uninfected G. roeseli individuals (Table 2, non-parametric multiple comparison analysis between groups corrected for tied ranks [43]). However, infected G. pulex individuals were far more photophilic than uninfected ones (Fig. 1). G. roeseli individuals (either infected or not) were significantly less photophobic than uninfected G. pulex.

In the Ouche River (Parc de la Colombière), the prevalence of P. laevis did not differ between the resident G. pulex and the invading G. roeseli. Our results somehow contrast with those obtained by Dunn and Dick [17] who found in an Irish river that the prevalence of the acanthocephalan Polymorphus minutus was significantly higher in the native G. duebeni celticus than in the invader G. tigrinus. However, a marked difference in prevalence of *P. laevis* (*G.* pulex: 11.56% (n = 450), G. roeseli: 2.69% (n = 253)) has been observed in a population (Les Maillys) distant from about 30 km from Parc de la Colombière (A. Bauer, unpublished results). More importantly, our results from the experiments on reaction to light suggest that the influence of P. laevis was actually lower in the invading host species than in the resident one. Infected G. roeseli individuals showed no altered behaviour in reaction to light compared to uninfected individuals of the same species, whereas infected G. pulex individuals were markedly less photophobic than uninfected ones. Although our results must be treated with caution since they are only based on one population, they contrast with a previous observation [44]. In this study, *Pomphorhynchus laevis* induced in the amphipod *Echinogammarus stammeri* the same behaviour as in *G*. pulex, characterized by a significant preference for lighted areas. However, Maynard et al. [44], unlike the present study, did not compare directly the effect of P. laevis on two host species coexisting in sympatry.

Because we used amphipods with natural infections in our experiments, there is a possibility that the observed modified behaviours lead to increased likelihood of infection, rather than the converse. However, previous studies of similar host-parasite systems (e.g. Bethel and Holmes [45])

Table 2
Results from multiple comparison between groups for reaction to light

Comparison	Observed <i>Q</i> value	P
Uninfected G. roeseli vs infected G. roeseli Uninfected G. roeseli vs uninfected G. pulex Uninfected G. roeseli vs infected G. pulex Infected G. roeselivs uninfected G. pulex Infected G. roeselivs infected G. pulex Uninfected G. pulexvs infected G. pulex	3.6654 - 3.9760 3.0741 - 3.9319 - 6.7270 0.3757	> 0.500 < 0.002 < 0.001 < 0.020 < 0.001 < 0.001

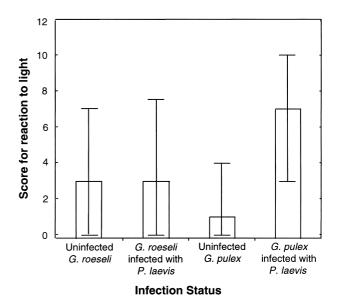


Fig. 1. Median values for reaction to light according to infection status. Bars show the interquartile range.

have shown that modified behaviours are observed only after the cystacanths have become infective to the definitive host, indicating that the observed alterations of infected hosts behavior are the consequence and not the cause of infection. We thus consider that the differential influence of the parasite between host species is the result of the interaction between the parasite's ability to manipulate its hosts and the ability of the hosts to resist manipulation [46,47]. We thus consider that the observed variation between host species in the virulence of the parasite is the result of the interaction between the parasite's ability to manipulate its hosts and the ability of the hosts to resist manipulation [46,47]. Therefore, both aspects of the interaction must be considered. Firstly, host specificity by the parasite might well account for the results obtained in this study. Although cases of infection of G. roeseli by P. laevis have been reported in eastern Europe [31,32] there is, to our knowledge, no detailed account of the influence of the acanthocephalan parasite on the behaviour of its host in this part of its geographic range. One possibility is that P. laevis is able to infest G. roeseli but is unable to alter its photophobic behaviour. The absence of enhanced photophily in G. roeseli infected with P. laevis in Burgundy may correspond to an adaptation of P. laevis to the predominant host species that might to some extent preclude its adaptation to the invading gammarid species. This would be in accordance with a recent theoretical study predicting that in a heterogeneous host population (composed of two host types) specialist parasite strains will evolve if the switch from one host to the other is connected with a high cost for the parasite [48]. Similarly, parasite-induced alterations in behaviour in insects are dependent [49,50] upon the species or even the genetical strain considered [50].

Finally, the inability of *P. laevis* to alter the behaviour of

G. roeseli might also be the consequence of the invasion process. According to Hynes [51], parasitised gammarids are more likely to die under stress than uninfected ones. If the colonisation of a new habitat involves a stress, then the invasion process may have selected for more vigorous individuals, or individuals resistant to acanthocephalan parasites [17]. Dobson and May [19] have indeed suggested that such a loss of parasites may confer a competitive advantage to the invading species. Comparative data on the behavioural effect of the parasite on G. roeseli in other areas of the geographic range and reciprocal cross-infection experiments are therefore necessary to determine whether resistant individuals were selected during the recent extension of the geographic range of G. roeseli, or if different strains of parasites are involved in the observed differential susceptibility of the two gammarid species to the parasite, as Munro et al. [52] mentioned in the British Isles.

Our results also pointed out, however, that *G. roeseli*, whether infected or not, were slightly less photophobic than healthy *G. pulex*. A change in behaviour induced by the parasite might thus be of little value, if the normal behaviour of *G. roeseli* already predisposes it to predation by an appropriate final host of *P. laevis*. However, *G. roeseli* clearly remained photophobic, although less so than *G. pulex*, and thus should be less exposed to predation than infected *G. pulex*. Direct experiments comparing the susceptibility of each gammarid species, whether infected or not by *P. laevis*, to predation by fish should help to clarify this point.

The overall study seems to indicate that these gammarid species, which share the same trophic niche and which have been recently found in sympatry, might be involved in an 'apparent' competition [18] potentially mediated via *P. laevis*. The absence of photophilic behavioural alteration in infected *G. roeseli* suggests less intense predation on the invading gammarid than on the resident one. This may have consequences on the population dynamics of the two species in the study area. Thus, long-term survey of both parasites and hosts should be undertaken to determine the role that parasites can play in regulating the success of invading species and hence shaping the structure of amphipod communities.

Acknowledgements

Financial support was provided by the Programme National Diversité Biologique, volet Interactions Durables and the Contrat de Plan Etat-Région Bourgogne. We thank Jérôme Comte, Marilyne Duval and Laurent Landré for field and laboratory assistance.

References

 Jazdzewski K. Range extension of some gammaridean species in European inland waters caused by human activity. Crustaceana 1980;S6:84–107.

- [2] Costello MJ. Biogeography of alien amphipods occurring in Ireland and interactions with native species. Crustaceana 1993;65:287–99.
- [3] Dick JTA. Post-invasion amphipod communities of Lough Neagh, Northern Ireland: influences on habitat selection and mutual predation. J Anim Ecol 1996;65:756–67.
- [4] van den Brink FWB, van der Velde G, Bij de Vaate A. Ecological aspects, explosive range extension and impact of a mass invader, *Corophium curvispinum*Sars, 1895 (Crustacea: Amphipoda), in the Lower Rhine (The Netherlands). Oecologia 1993;93:224–32.
- [5] Dick JTA, Platvoet D. Invading predatory crustacean *Dikerogam-marus villosus* eliminates both native and exotic species. Proc R Soc Lond B 2000;267:977–83.
- [6] Dennert HG. Tolerance differences and interspecific competition in three members of the amphipod genus *Gammarus*. Bijdr Dierkun 1974:44:83–89.
- [7] Dick JTA, Elwood RW. Coexistence and exclusion among *Gammarus* species: behavioural avoidance of interspecific precopulation by male *G. pulex* (Amphipoda). Oikos 1992;64:541–7.
- [8] Dick JTA, Montgomery WI, Elwood RW. Replacement of the indigenous amphipod *Gammarus duebeni celticus* by the introduced *G. pulex*: differential cannibalism and mutual predation. J Anim Ecol 1993;62:79–88.
- [9] Dick JTA, Elwood RW, Montgomery WI. The behavioural basis of a species replacement: differential aggression and predation between the introduced *Gammarus pulex* and *G. duebeni celticus* (Amphipoda). Behav Ecol Soc 1995;37:393–8.
- [10] Freeland WJ. Parasites and the coexistence of animal host species. Am Nat 1983;121:223–36.
- [11] Holt RD, Pickering J. Infectious diseases and species coexistence: a model of Lotka-Volterra form. Am Nat 1985;126:196–211.
- [12] Price PW, Westoby M, Rice B. Parasite-mediated competition: some predictions and tests. Am Nat 1988;131:544–55.
- [13] Park T. Experimental studies of competition. I. Competition between populations of the flour beetles, *Tribolium confusum* Duval and *Tribolium castaneum* Herbst. Ecol Monog 1948;18:265–308.
- [14] Price PW, Westoby M, Rice B, et al. Parasite mediation in ecological interactions. Ann Rev Ecol Syst 1986;17:487–505.
- [15] Richner H, Oppliger A, Christe P. Effects of an ectoparasite on the reproduction in great tits. J Anim Ecol 1993;62:703–10.
- [16] Thomas F, Renaud F, Rousset F, Cézilly F, De Meeüs T. Differential mortality of two closely related host species induced by one parasite. Proc R Soc Lond B 1995;260:349–52.
- [17] Dunn AM, Dick JTA. Parasitism and epibiosis in native and nonnative gammarids in freshwater in Ireland. Ecography 1998;21:593– 8.
- [18] Hudson P, Greenman J. Competition mediated by parasites: biological and theoretical progress. Trends ecol evol 1998;13:387–90.
- [19] Dobson AP, May RM. Patterns of invasion by pathogens and parasites. In: Mooney HA, Drake JA, editors. Ecology of biological invasions in North America and Hawaii, 58. Ecol Stud 1986. pp. 58–76.
- [20] Holdich DM, Reeve ID. Distribution of freshwater crayfish in the British Isles, with particular reference to crayfish plague, alien introductions and water quality. Aq Cons 1991;1:139–58.
- [21] Settle WH, Wilson LT. Invasion by the variegated leafhopper and biotic interactions: parasitism, competition and apparent competition. Ecology 1990;71:1461–70.
- [22] Crompton DWT, Nickol BB. Biology of the Acanthocephala. Cambridge: Cambridge University Press, 1985.
- [23] Moore J, Gotelli NJ. Phylogenetic perspective on the evolution of altered host behaviours: a critical look at the manipulation hypothesis. In: Barnard CJ, Behnke JM, editors. Parasitism and host behaviour, 1990. pp. 193–233.
- [24] Combes C. Ethological aspects of parasite transmission. Am Nat 1991;138:866–80.
- [25] Bethel WM, Holmes JC. Altered evasive behavior and responses to light in amphipods harboring acanthocephalan cystacanths. J Parasitol 1973;59:945–56.

- [26] Bakker TCM, Mazzi D, Zala S. Parasite-induced changes in behavior and color make *Gammarus pulex* more prone to fish predation. Ecology 1997;78:1098–104.
- [27] Cézilly F, Grégoire A, Bertin A. Conflict between co-occurring manipulative parasites? An experimental study of the joint influence of two acanthocephalan parasites on the behaviour of *Gammarus* pulex. Parasitology 2000;120:625–30.
- [28] Karaman GS, Pinkster S. Freshwater Gammarus species from Europe, North Africa and adjacent regions of Asia (Crustacea Amphipoda). II. Gammarus roeseli-group and related species. Bijdr Dierk 1977;47:165–96.
- [29] Roux AL, Roux C, Opdam Y. Répartition écologique et métabolisme respiratoire de *Gammarus roeseli* Gervais 1835. Crustaceana 1980;S6:148–59.
- [30] Jazdzewski K, Roux AL. Biogéographie de Gammarus roeseli Gervais en Europe, en particulier répartition en France et en Pologne. Crustaceana 1988;S13:272–7.
- [31] Rašin K. Contributions to the biology of the species Pomphorhynchus laevis Müll. (= Echinorhynchus proteus Westrumb). Zpravodaj Moraského zemského rybášského spolku v Brné, Brno 1928:44–64.
- [32] Moravec F, Scholz T. Observations on the biology of *Pomphor-hynchus laevis* (Zoega in Müller, 1776) (Acanthocephala) in the Rokytna River, Czech and Slovak Federative Republic. Helminthologia 1991;28:23–29.
- [33] Hine PM, Kennedy CR. Observations on the distribution, specificity and pathogenicity of the acanthocephalan *Pomphorhynchus laevis* (Müller). J Fish Biol 1974;6:521–35.
- [34] Rumpus AE, Kennedy CR. The effect of the acanthocephalan Pomphorhynchus laevis upon the respiration of its intermediate host Gammarus pulex. Parasitology 1974;68:271–84.
- [35] Bentley CR, Hurd H. Pomphorhynchus laevis (Acanthocephala): elevation of haemolymph protein concentrations in the intermediate host Gammarus pulex (Crustacea: Amphipoda). Parasitology 1993;107:193–8.
- [36] Bentley CR, Hurd H. Carbohydrate titres in the haemolymph and midgut glands of *Gammarus pulex* infected with the acanthocephalan *Pomphorhynchus laevis*. J Helminthol 1996;70:103–7.
- [37] Kennedy CR, Broughton PF, Hine PM. The status of brown and rainbow trout, Salmo trutta and S. gairdneri as hosts of the acanthocephalan Pomphorhynchus laevis. J Fish Biol 1978;13:265–75.

- [38] Hynes HBN. The reproductive cycle of some British freshwater Gammaridae. J Anim Ecol 1955;24:352–87.
- [39] Bollache L, Gambade G, Cézilly F. The influence of micro-habitat segregation on size assortative pairing in *Gammarus pulex* (L.) (Crustacea, Amphipoda). Arch Hydrobiol 2000;147:547–58.
- [40] Brauer A. Die Süsswasserfauna Deutschlands. Heft 16: Acanthocephalen. Jena: Verlag von Gustav Fischer, 1911.
- [41] Brown AF, Chubb JC, Veltkamp CJ. A key to the species of Acanthocephala parasitic in British freshwater fishes. J Fish Biol 1986;28:327–34.
- [42] Siegel S, Castellan NJ. Nonparametric statistics for behavioral sciences. Second edition. New York, NY: Mc Graw-Hill, 1988.
- [43] Zar JH. Biostatistical analysis, 3rd ed. London: Prentice-Hall Limited Editions, 1996.
- [44] Maynard BJ, Wellnitz TA, Zanini N, Wright WG, Dezfuli BS. Parasite-altered behavior in a crustacean intermediate host: field and laboratory studies. J Parasitol 1998;84:1102–6.
- [45] Bethel WM, Holmes JC. Correlation of development of altered evasive behavior in *Gammarus lacustris* (Amphipoda) harboring cystacanths of *Polymorphus paradoxus* (Acanthocephala) with the infectivity to the definitive host. J Parasitol 1974;60:272–4.
- [46] Poulin R. Evolutionary Ecology of parasites. From individuals to communities. London: Chapman & Hall, 1998.
- [47] Poulin R, Combes C. The concept of virulence: Interpretations and Implications. Parasitol Today 1999;15:474–5.
- [48] Regoes RR, Nowak MA, Bonhoeffer S. Evolution of virulence in a heterogeneous host population. Evolution 2000;54:64–71.
- [49] Moore J, Gotelli NJ. Evolutionary patterns of altered behavior and susceptibility in parasitized hosts. Evolution 1996;50:807–19.
- [50] Yan G, Stevens L, Schall JJ. Behavioural changes in *Tribolium* beetles infected with a tapeworm: variation in effects between beetle species and among genetic strains. Am Nat 1994;143:830–47.
- [51] Hynes HBN. The ecology of Gammarus duebeni Lilljeborg and its occurrence in fresh water in western Britain. J Anim Ecol 1954:23:38–84.
- [52] Munro MA, Reid A, Whitfield PJ. Genomic divergence in the ecologically differentiated english freshwater and marine strains of *Pomphorhynchus laevis* (Acanthocephala: Palaeacanthocephala): a preliminary investigation. Parasitology 1990;101:451–4.